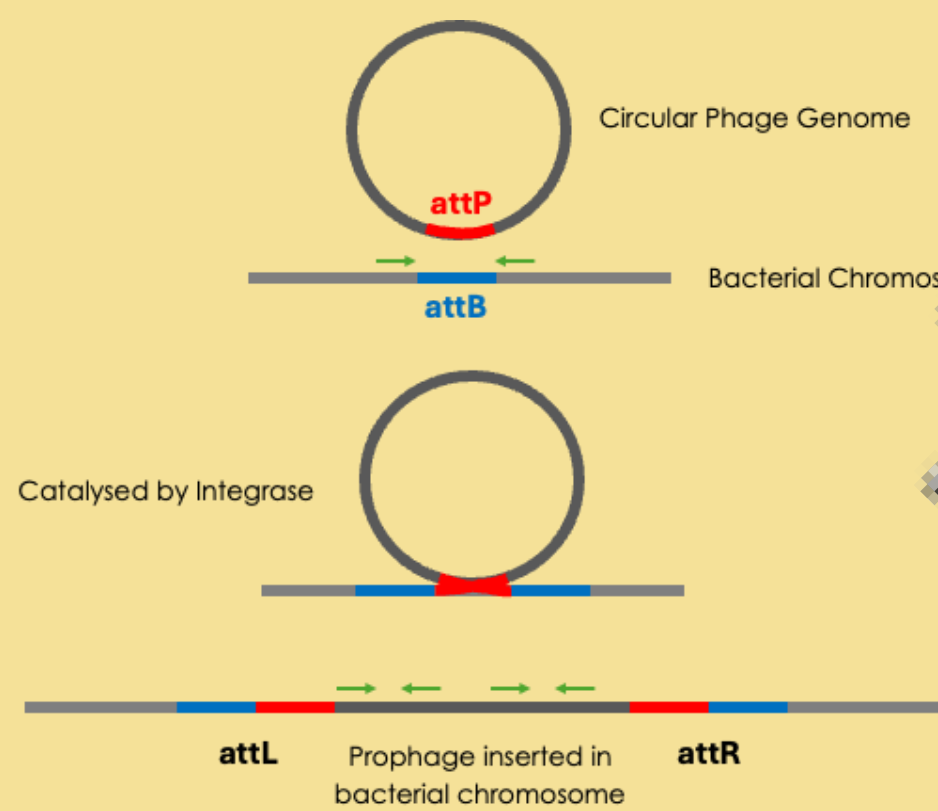


# Targeted engineering of phages to understand infection mechanisms

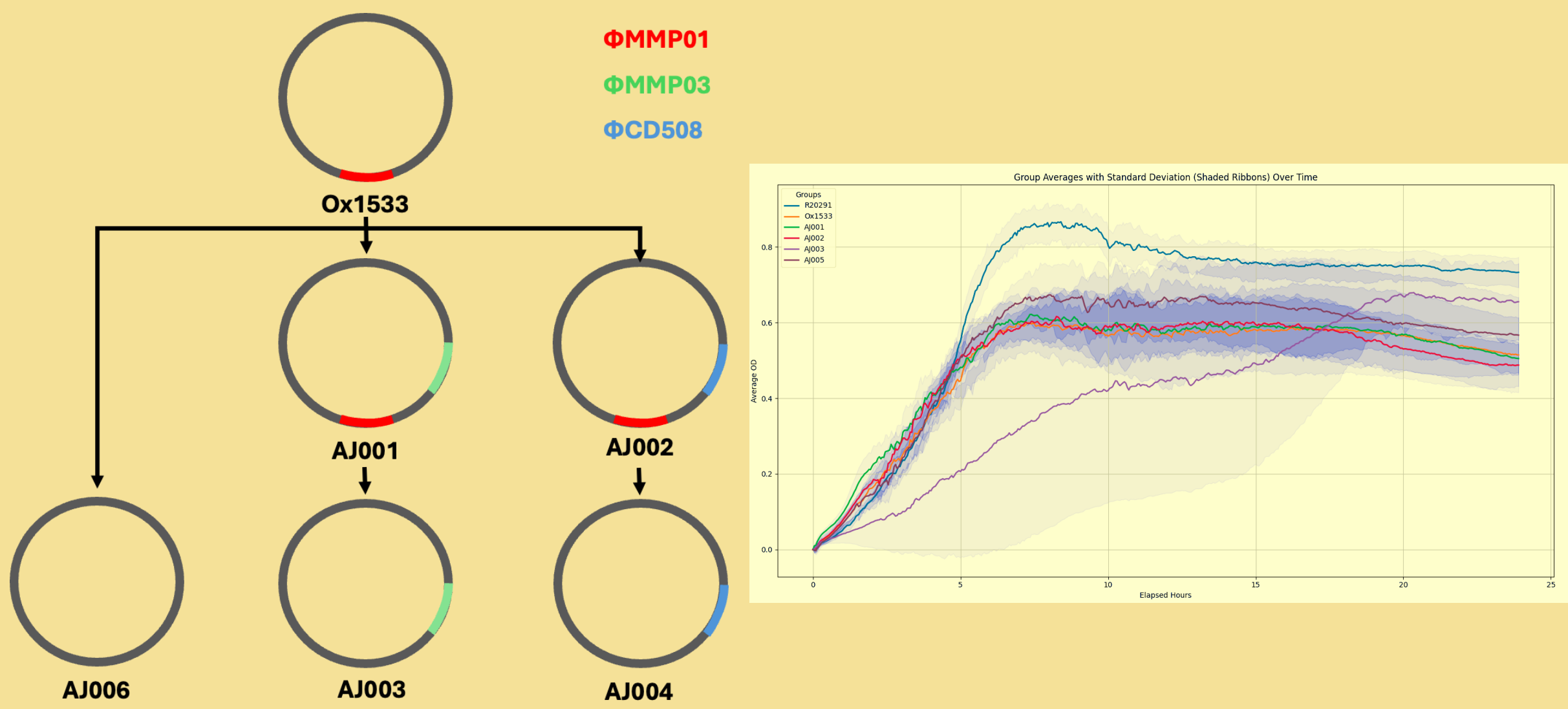
Anirudh Jakhmola,<sup>1</sup> Jason S. Wilson,<sup>1</sup> Rebbekah Menda,<sup>1</sup> Louis Charles Fortier,<sup>2</sup> Per Bullough,<sup>1</sup> Robert P. Fagan<sup>1</sup>  
<sup>1</sup>University of Sheffield, United Kingdom; <sup>2</sup>Université de Sherbrooke, Canada

## Introduction

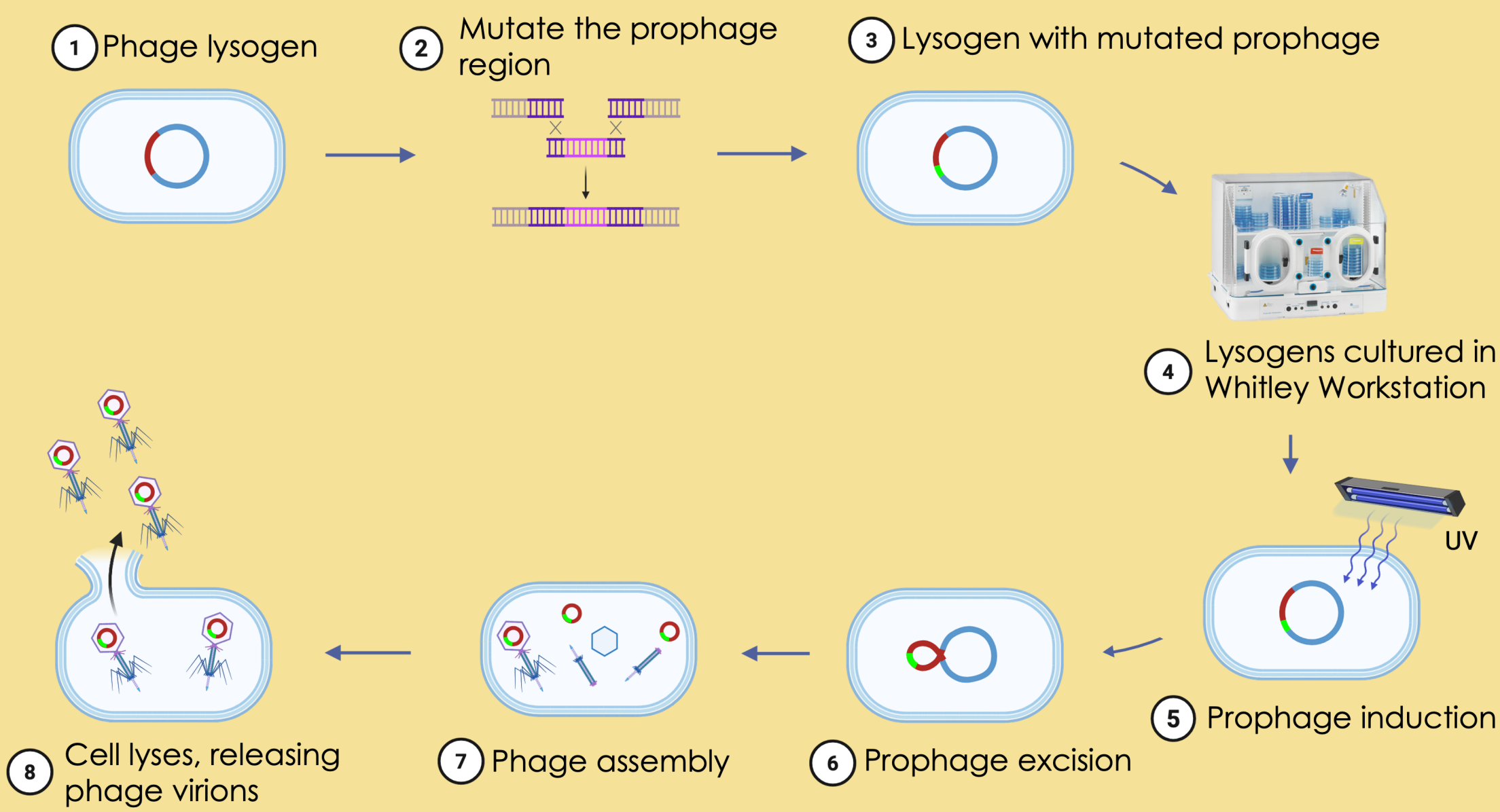
- There has been a lack of *C. difficile* phage studies due to their difficult molecular biology.
- We made *C. difficile* lysogens in order to be able to mutate them.
- Using the structure of phages, we discovered two enzymes on the needle tip of a phage that potentially help the phage in infection.
- Phages and bacteria were cultured in Whitley DG250 Workstation to ensure optimum growth conditions for infections.



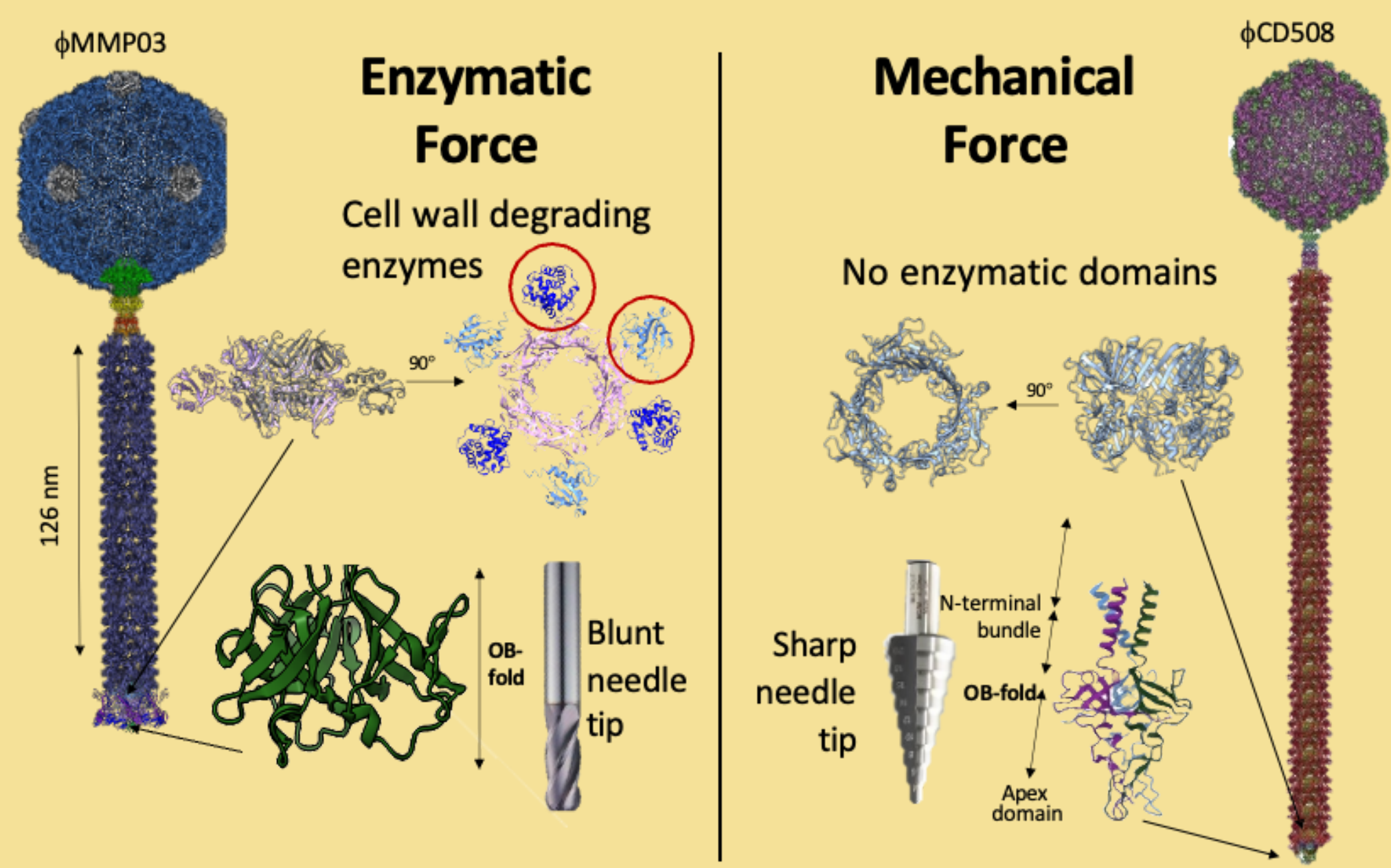
## Strain Background



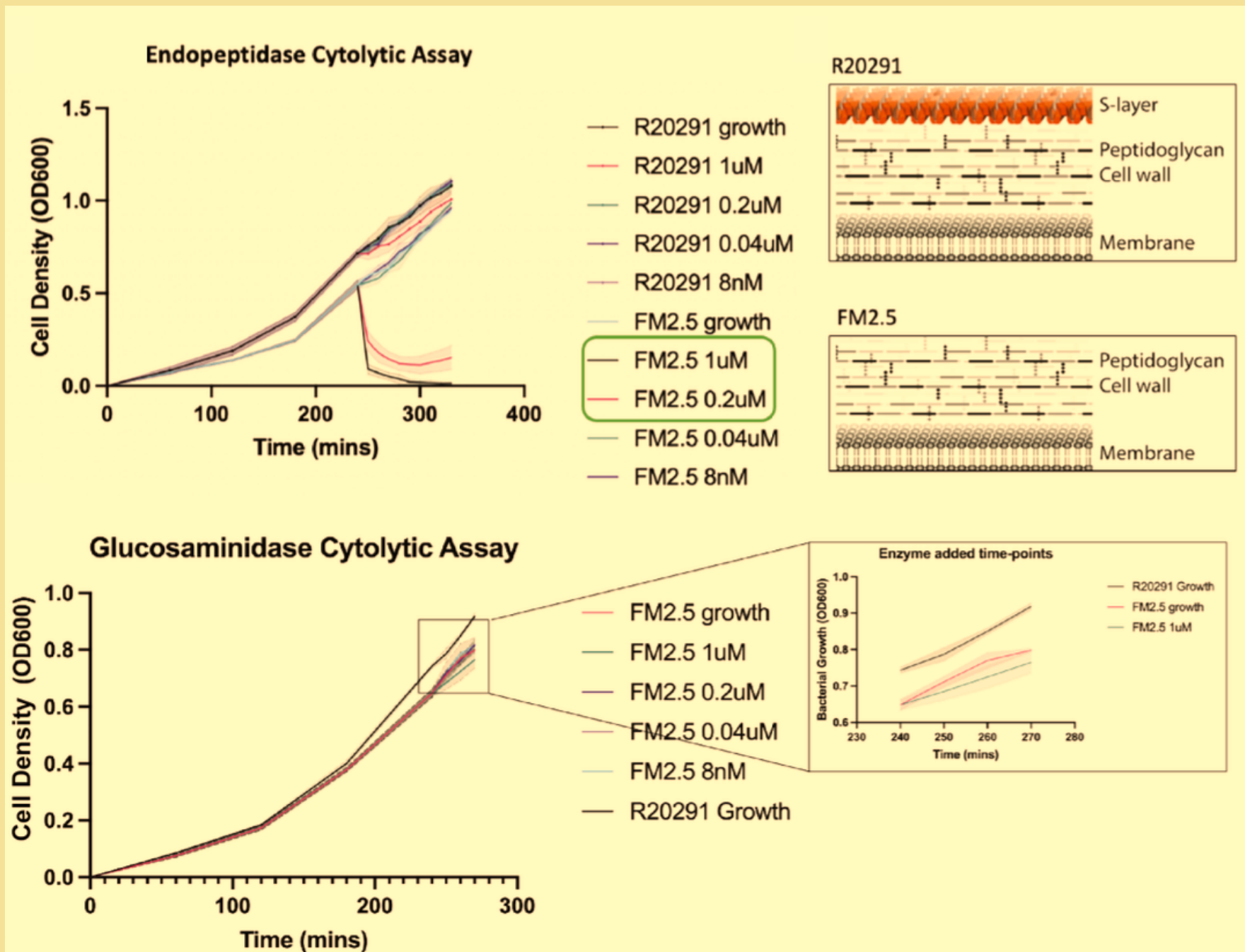
## Phage Mutagenesis Pipeline



## Difference Between Needle Tips

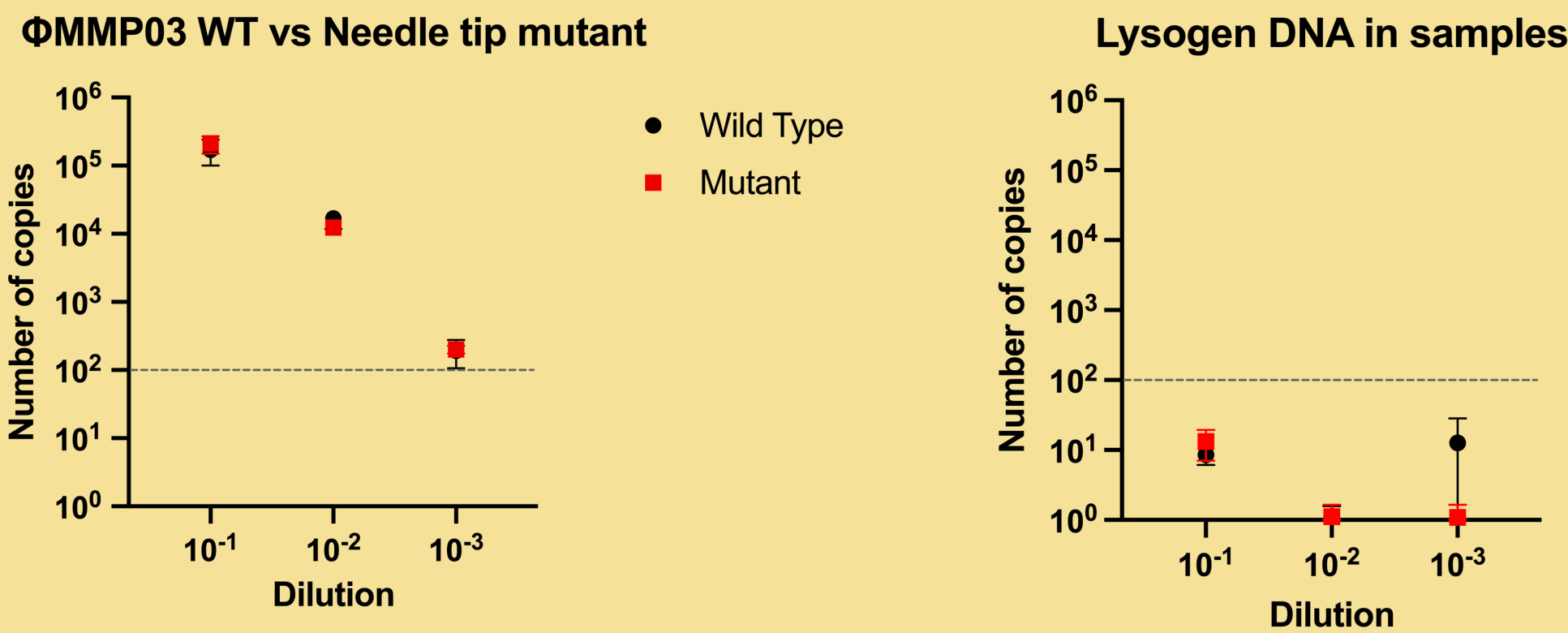


## In Vitro Cytolytic Assays

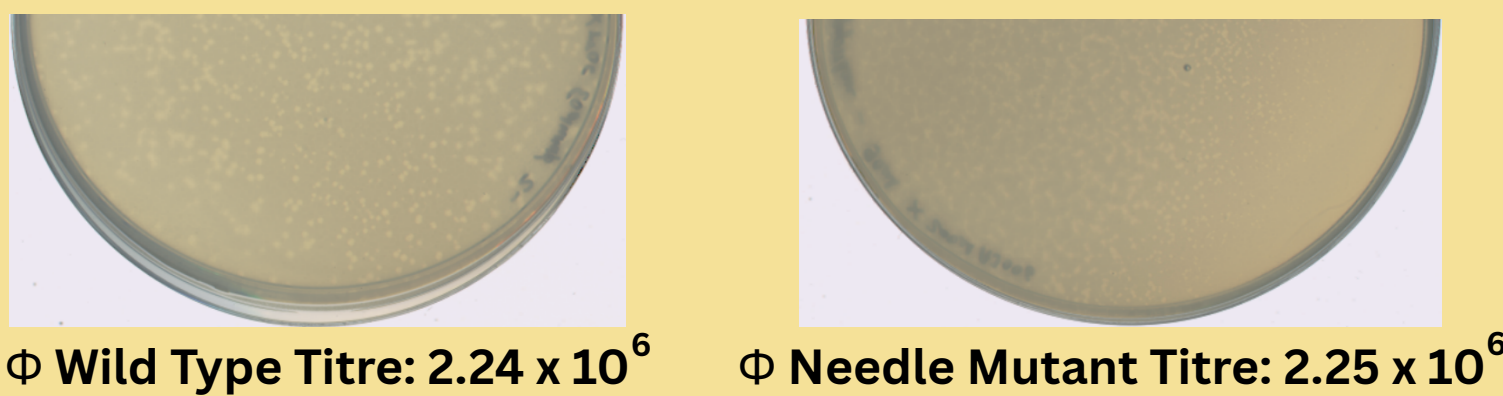


## Needle Tip Mutant Assays

### qPCR assay



### Infection assay



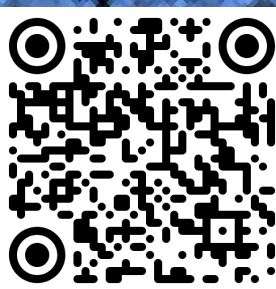
The results showed no significant difference between the wild type and the mutant, suggesting a potential role of glucosaminidase in infection.

## Next Questions

- Does glucosaminidase play a more important role in infection?
- Is there any interaction between endopeptidase and glucosaminidase that affects the infection efficiency?
- If not, what is the evolutionary significance of the enzymatic domains in the tip?

### Next steps:

- Apply the mutagenesis method to answer if tail length and level of contraction of the phage affect infection efficiency.
- Make phages modular in order to use them as a tool in *C. difficile* research.



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